

Prenatal Screening for Fetal Aneuploidy

Policy Number: AHS– G2055 – Prenatal Screening for Fetal Aneuploidy	Prior Policy Name and Number, as applicable: <ul style="list-style-type: none"> • AHS-G2055-20161212-Detection of Down Syndrome Using Maternal Serum Markers • AHS-M2016-20170320-Non-Invasive Prenatal Screening for Aneuploidy
Initial Presentation Date: Revision Date: N/A	
Policy outlines coverage for Arkansas PASSE	

I. Policy Description

Aneuploidy is defined as an abnormal number of chromosomes present in the cell. Fetal aneuploidy is a condition where the fetus has one or more extra or missing chromosomes leading to either a nonviable pregnancy, offspring that may not survive after birth, or surviving newborn with congenital birth defects and functional abnormalities. The most common fetal aneuploidies associated with an additional chromosome are Down syndrome (trisomy 21), Edwards syndrome (trisomy 18), and Patau syndrome (trisomy 13). Prenatal screening for fetal aneuploidy is an assessment of the woman’s risk of carrying a fetus with fetal aneuploidy using markers found in maternal serum (ACOG, 2016). Non-invasive prenatal screening is a method for screening for chromosomal abnormalities using a maternal blood sample where cell-free fetal DNA (cff-DNA) is extracted and screened for aneuploidies (McKanna et al., 2018).

II. Related Policies

Policy Number	Policy Title
AHS-G2035	Prenatal Screening
AHS-M2033	Chromosomal Microarray

III. Indications and/or Limitations of Coverage

Medical Policy Statements do not ensure an authorization or payment of services. Please refer to the plan contract (often referred to as the Evidence of Coverage) for the service(s) referenced in the Medical Policy Statement. If there is a conflict between the Medical Policy Statement and the plan contract (i.e., Evidence of Coverage), then the plan contract (i.e., Evidence of Coverage) will be the controlling document used to make the determination.

Application of coverage criteria is dependent upon an individual's benefit coverage at the time of the request. If there is a conflict between this Policy and any relevant, applicable government policy [e.g. National Coverage Determinations (NCDs) for Medicare] for a particular member, then the government policy will be used to make the determination. For the most up-to-date Medicare policies and coverage, please visit their search website <http://www.cms.gov/medicare-coverage-database/overview-and-quick-search.aspx?from2=search1.asp&> or the manual website

Application of coverage criteria is dependent upon an individual's benefit coverage at the time of the request

1. Screening test to detect Fetal Aneuploidy of chromosomes 13, 18 and 21 **MEET COVERAGE CRITERIA** for women who are adequately counseled and desire information on the risk of having a child with Fetal Aneuploidy (e.g. Down syndrome) under following conditions:
 - a. First-trimester (defined as 11-14 weeks) screening incorporating maternal serum markers (hCG, PAPP-A with NT).
 - b. Second-trimester (15-22 weeks) screening incorporating triple maternal serum markers (hCG, AFP, uE3 with NT) & Quad maternal serum markers (hCG, AFP, uE3, DIA with NT).
 - c. First (11-14 weeks) & second (15-22 weeks) trimester integrated screening incorporating maternal serum markers (PAPP-A with NT) & Quad maternal serum markers (hCG, AFP, uE3, DIA with NT).
 - d. First (11-14 weeks) & second (15-22 weeks) trimester sequential screening incorporating maternal serum markers (PAPP-A, hCG with NT) & Quad maternal serum markers (hCG, AFP, uE3, DIA with NT).
 - e. First (11-14 weeks) & second (15-22 weeks) trimester contingent screening incorporating maternal serum markers (PAPP-A, hCG with NT) if positive, Quad maternal serum markers (hCG, AFP, uE3, DIA with NT).
 - f. First & second trimester non-invasive prenatal screening (NIPS) for fetal aneuploidy (of at least 10 weeks gestation and singleton pregnancy) incorporating maternal serum cell-free fetal DNA.
2. Confirmatory testing of equivocal and positive results from testing listed above via Chorionic Villa Sampling (CVS) or Amniocentesis should be offered and **MEETS COVERAGE CRITERIA** for women wishing to pursue additional testing.

3. The use of the “penta” screen (hCG, AFP, uE3, DIA with NT, and hyperglycosylated hCG) to detect fetal aneuploidy **DOES NOT MEET COVERAGE CRITERIA.**
4. Screening for detection of Fetal Aneuploidies **DOES NOT MEET COVERAGE CRITERIA** under following conditions:
 - a. Parallel or simultaneous testing with multiple screening methodologies for Fetal aneuploidy.
 - b. Screening of women with multiple gestation pregnancies with any testing other than nuchal translucency and/or subsequent diagnostic testing via Chorionic Villa Sampling (CVS) or Amniocentesis due to the risk of high false positive results.
 - c. Repeat screening for women with negative screening results.
 - d. Egg donor pregnancies
 - e. For all uses other than the detection of fetal trisomy of 13, 18, and 21 and Turner syndrome (e.g. Microdeletion syndromes, unbalanced translocations, deletions, duplications).
 - f. For the determination of fetal sex

IV. Scientific Background

Pregnant women are routinely offered blood-based screening or invasive diagnostic testing for identification of the most common fetal aneuploidies: trisomy 13 (Patau syndrome), trisomy 18 (Edwards syndrome) and trisomy 21 (Down syndrome). Approximately 90% of chromosomal abnormalities are due to an incorrect number of chromosomes, especially in the three aforementioned triploid conditions as well as monosomy X (Turner syndrome). Approximately 15-20% of clinically recognized pregnancies result in first trimester spontaneous abortions, with 50% of these spontaneous abortions due to chromosomal abnormalities (Witters et al., 2011).

Historically, non-invasive blood-based aneuploidy screening has taken the form of first- and/or second-trimester analysis of biomarkers in maternal circulation, sometimes along with ultrasound measurement of fetal nuchal translucency (NT). Although both the sensitivity (detection rate) and specificity (true positive rate) of maternal serum screening tests for aneuploidy have improved significantly over time, the false positive rate (2-5%) remains higher than desirable. The detection rate of Down syndrome in the first trimester using a combination of NT and biochemical markers is typically 79% - 90% (Dey, Sharma, & Aggarwal, 2013). Positive maternal serum screen results are usually followed by an invasive diagnostic test, such as karyotyping of a chorionic villus sample (in first trimester) or karyotyping of an amniotic fluid sample (second trimester).

Additionally, detection rates of maternal serum screens are typically below 99%, resulting in the inability of a normal result to confer complete confidence that the fetus is unaffected with aneuploidy (Dey et al., 2013). Thus, many women who are in a high-risk category due to age or other factors may opt for the more definitive, diagnostic, invasive testing, which has its own risks and relatively high costs. The availability of non-invasive testing may improve both the sensitivity and specificity of aneuploidy detection while resulting in fewer invasive procedures, less risk, and less overall cost.

Screening Tests

Chromosomal anomalies are a leading cause of perinatal mortality and developmental abnormality. The goal of prenatal testing is to screen for chromosomal anomalies and to provide genetic counseling for parents. The American College of Obstetrics and Gynecology (ACOG) recommends that prenatal testing is offered to all pregnant women (ACOG, 2016, 2020). Invasive testing, including chorionic villi sampling or amniocentesis, should be limited to high-risk patients owing to the potential risks for procedure-related pregnancy loss.

Genome-wide sequencing tests for fetal diagnostics have also been employed and are expected to increase in popularity as the cost decreases and as new tools are developed. These tests include DNA sequencing methods, such as whole exome-sequencing and targeted clinical panels, which can further evaluate fetal structural anomalies first detected in an ultrasound (ISPD, 2018). This diagnostic sequencing method has been used for various fetal diagnostic measures including standard genetic testing and chromosomal microarray analysis.

Chromosomal microarray (CMA) testing refers to the use of comparative genomic hybridization (CGH) arrays to compare the DNA of a patient with a normal control (Aradhya, Manning, Splendore, & Cherry, 2007). CMA is significantly more sensitive (10 to 100 kb) than traditional karyotyping (5 to 10 Mb), has a turnaround time of five days quicker than karyotyping (Robson et al., 2017), and provides an alternative to karyotyping when dividing cells are not available for analysis. This technique may be used for several different purposes, such as identifying a cause of pregnancy loss or identifying other aneuploid conditions, such as Down Syndrome (Reddy et al., 2012). This method of diagnostic prenatal sequencing is currently investigational because of limited data and is utilized most prominently in research settings or clinically on a case-by-case basis (ISPD, 2018).

Biochemical Markers in Maternal Serum

Many studies revealed that maternal age, fetal NT, maternal serum free β -human chorionic gonadotropin (hCG) and pregnancy-associated plasma protein-A (PAPP-A) have been associated with aneuploidy. The "Quad screen," comprising alpha-fetoprotein (AFP), hCG, unconjugated estriol (E3), and inhibin-A, is the most efficient multiple-marker screening test in the second trimester. In addition, there are more options such as integrated, sequential testing, and cell-free DNA screening. Many studies are ongoing to reveal the most sensitive, specific and effective screening tools for use during the first trimester (Park et al., 2016).

To improve the accuracy of serum markers, ultrasound markers are used. NT refers to the fluid filled space measured on the dorsal aspect of the fetal neck. An enlarged NT (>3.0 mm/99th percentile of the crown-rump length) is independently associated with fetal aneuploidy and structural malformations (ACOG, 2016, 2020).

Screening studies of pregnant women reported an association between increased NT in the first trimester of pregnancy (10-13 weeks of gestation) and chromosomal defects, most commonly Down syndrome (trisomy 21) but also trisomy 18 and 13. NT could be done alone as a first-trimester screen or in combination with maternal serum markers, free beta subunit of human chorionic gonadotropin (β -hCG) and pregnancy-associated plasma protein-A (PAPP-A). All three trisomies (chromosomes 13, 18, and 21) "are associated with increased maternal age, increased fetal NT and decreased PAPP-A, but in trisomy 21 serum free β -hCG is increased whereas in trisomies 18 and 13 free β -hCG is

decreased” (Shiefa, Amargandhi, Bhupendra, Moulali, & Kristine, 2013). Low β -hCG in the first trimester has also been associated with an increased risk of significant copy number variants on chromosomal microarray analyses (Bornstein et al., 2018).

Analytical Validity of Biochemical Markers

Screening for chromosomal abnormalities using biochemical markers include the first trimester combined test, triple test, quadruple test, sequential test, and integrated test. Except for the first trimester combined test, all others can provide screening results in the second trimester. In the first trimester combined test, the risk is calculated based on the ultrasonographic findings of NT and maternal serum levels of free β -hCG and PAPP-A. First-trimester screening not only allows early reassurance or early diagnosis of aneuploidy, but it also provides an option of earlier and safer termination of pregnancy in affected cases. Consequently, the first trimester combined test has become one of the most popular and useful screening strategies. Lee, Chen, Cheong, Chou, and Tsai (2013) conducted a 13-year study of 25,104 pregnant women using the first trimester Down syndrome screening. “The detection rates for trisomy 21, trisomy 18, Turner syndrome, and other chromosome anomalies were 87.5% (21/24), 69.2% (9/13), 81.8% (9/11), and 60% (18/30), respectively, with a false positive rate (FPR) of 5.4% (1353/25,026). Further evaluation of the detection rates for trisomy 21, by gestational age at 11, 12, and 13 weeks, were 92.3%, 87.5%, and 66.7%, respectively” (Lee et al., 2013).

For second-trimester screening for Down syndrome, the sensitivity and specificity of the triple test—co-testing AFP, unconjugated E3, and free β -hCG—are higher than screening with AFP alone. However, when the false-positive rate is fixed at 5% in order to compare the screening performance between the screening tools, the detection rate was found to be 66.8% to 77% with the triple test and 75.9% to 92% with the first trimester combined test. The sensitivity of the triple test was lower than the combined test (Baer et al., 2015).

The quadruple test, which uses the fourth marker, inhibin-A, in addition to the other three markers, has 7% higher sensitivity when applying a fixed 5% false-positive rate. A study conducted by Wald et al. (2003) revealed that when inhibin-A was added to the traditional triple marker test, a detection rate of 83% was achieved, which was 6% higher than the 77% detection rate found with the triple test. This result was similar to that produced with the first trimester combined test (Park et al., 2016).

Many studies, including the Serum, Urine and Ultrasound Screening Study (SURUSS) (Wald et al., 2003) and the First-and Second-Trimester Evaluation of Risk (FASTER) study (Malone et al., 2005) have offered evidence suggesting that first-trimester screening for Down syndrome with measurement of fetal NT and maternal serum markers is at least as accurate as alternative tests and may allow earlier confirmation or exclusion of Down syndrome. These studies evaluated several tests in parallel, including first-trimester testing with NT and maternal markers, the triple test, second-semester quadruple test and a combined first- and second-trimester test (both with and without NT), stepwise sequential testing (results given after first-trimester testing, move on to second-trimester testing), and integrated screening (results given only after first and second-trimester testing). In a direct comparison of the first-trimester test to the triple test, the SURUSS study has shown that setting the false-positive rate at 5% resulted in an 83% detection rate, which was superior to what was historically expected of the triple test (Wald et al., 2003). SURUSS results were based on data from 47,053 pregnancies (101 with Down syndrome). The FASTER trial was conducted in the United States and was sponsored by the National Institutes of Health. The study enrolled 38,167 women and provided

further evidence that first-trimester combined screening was effective, but it did not provide NT measurement alone; results showed that integrated first- and second-trimester screening provided higher detection rates. The SURUSS and FASTER studies also found that overall, first-trimester screening with NT alone is inferior to either first- or second-trimester combined screening. Additional testing may not be necessary in those few cases when NT is at least 4.0 mm due to the high likelihood of Down syndrome in these cases (Malone et al., 2005; Wald et al., 2009; Wald et al., 2003).

Studies have found a high rate of successful imaging of the fetal nasal bone and an association between absent nasal bone and the presence of Down syndrome in high-risk populations. However, there is insufficient evidence on the performance of fetal nasal bone assessment in average-risk populations. Of concern is the low performance of fetal nasal bone assessment in a subsample of the FASTER study conducted in a general population sample. Two studies conducted outside of the United States have found that, when added to a first-trimester screening program evaluating maternal serum markers and NT, fetal nasal bone assessment can result in a modest decrease in the false-positive rate. Several experts in the field are proposing that fetal nasal bone assessment be used as a second stage of screening to screen women found to be of borderline risk using maternal serum markers and NT. Considering the uncertainty of test performance in average-risk populations and the lack of standardization in the approach to incorporating this test into a first-trimester screening program, detection of fetal nasal bone is considered investigational (Wald et al., 2009).

Cell-Free Fetal DNA from Maternal Serum

In 1997, researchers reported the identification of cell-free fetal DNA (cff-DNA) in the circulation of pregnant women (Lo et al., 1997). The fetal fraction of cff-DNA increases throughout gestation. At 10 weeks of gestation, the fetal fraction comprises at least 3 – 4% whereas it can account for up to 50% of the total cell-free DNA at term (G. E. Palomaki, Messerlian, & Halliday, 2018). Since then, laboratories have validated several different techniques for the use of cell-free DNA as a screening test for fetal aneuploidy, and these methods have been termed Non-Invasive Prenatal Screening or Testing (NIPS or NIPT).

Non-Invasive Prenatal Screening is a testing method which utilizes cell-free DNA from the plasma of pregnant women to screen for fetal aneuploidy. It is important to note that cell-free DNA screening does not assess risk of fetal anomalies, including neural tube defects or ventral wall defects (ACOG, 2015). NIPS methods only provide an estimate of whether the risk of aneuploidy is increased or decreased; NIPS does not provide a definitive diagnosis of aneuploidy. As with other aneuploidy screening tests, it is recommended that positive results of NIPS be followed by diagnostic testing such as traditional karyotyping of fetal cells obtained via chorionic villus sampling or amniocentesis (Gregg et al., 2016).

One cell-free fetal DNA detection method for NIPS, known as massively parallel sequencing (MPS), is a technique in which millions of pieces of maternal and fetal chromosomal material are sequenced and quantified. The MPS method is able to detect many types of aneuploidies, including those which are less commonly seen (Devers et al., 2013). MPS can detect common aneuploidies with both high sensitivity and high specificity for trisomies 13, 18, and 21. Bianchi et al. (2012) found the detection rate sensitivity for trisomy 21 to be 100%, the detection rate sensitivity for trisomy 18 to be 97.2%, and the detection rate sensitivity for trisomy 13 to be 78.6%; specificity was 100% for all three of the aforementioned trisomies.

Detection of aneuploidy using circulating cell-free fetal DNA can also be performed using selective analysis of specific loci only from the chromosomes of interest, as opposed to sequencing of all chromosomes performed in MPS. This directed analysis of cell-free fetal DNA has also been shown to have high sensitivity and high specificity for the common trisomies. Lee et al. (2019) utilized plasma from 1,055 pregnant woman and found that NIPT with cell-free fetal DNA “showed 100% sensitivity and 99.9% specificity for trisomy 21, and 92.9% sensitivity and 100% specificity for trisomy 18, and 100% sensitivity and 99.9% specificity for trisomy 13.”

The third approach to detect aneuploidy from cff-DNA is based on the amplification of single nucleotide polymorphisms (SNPs) on the chromosome of interest. In a study by Eiben et al. (2015), 2,942 patients underwent SNP-based non-invasive prenatal screening (NIPS) in which the source for cff-DNA was derived from placental cells. Sixty-five patients (2.2%) had positive non-invasive prenatal screening results for aneuploidy and further invasive testing confirmed aneuploidy in fifty-nine of those patients (90.8%). The remaining six patients were false positives due to a discrepancy between the genetic status of the fetus and placenta, a condition known as confined placental mosaicism (CPM). The fetal fraction was abnormally low (<8%) and indicative of fetal-placental discrepancies. Although a reliable screening method, the author suggests that SNP-based NIPS “cannot be used as a standalone test without ultrasound examination or invasive confirmation (Eiben et al., 2015).”

Despite the apparent advantages of NIPS over standard maternal serum screening in screening for common aneuploidies, there are limitations. “Reported Ifs [incidental findings] range from fetal or maternal deletions and duplications or mosaic sex chromosome aneuploidy in the mother or fetus, presenting as aneuploidy risk on NIPS, to mosaicism and uniparental disomy to abnormal results because of the presence of cell-free DNA originating from an undiagnosed maternal tumor” (Westerfield, Darilek, & van den Veyver, 2014). When ultrasound evaluation reveals fetal anomalies that may be consistent with one of those scenarios, invasive diagnostic testing with karyotyping or microarray may be more appropriate. NIPS also cannot distinguish the cause of aneuploidy, nor can it differentiate among the presence of an extra chromosome, a Robertsonian translocation, or high-level mosaicism. The determination of the type of aneuploidy is important for accurate counseling and future risk assessment (Neufeld-Kaiser, Cheng, & Liu, 2015; Strom et al., 2017; Westerfield et al., 2014). Also, some samples contain insufficient amounts of cell-free DNA, which is unknown until the test procedure has commenced. Early gestational age (<10 weeks) and high body mass index have been shown to be associated with reduced amounts of circulating cell-free fetal DNA. Additionally, NIPS for aneuploidy does not detect the presence of neural tube defects, which is included in traditional second trimester maternal serum screening. It has been suggested that the testing of maternal serum AFP in the second trimester should be offered to women who underwent first-trimester aneuploidy screenings (G. E. Palomaki, Messerlian, & Halliday, 2020).

Analytical Validity of Cell-free Fetal DNA Testing

A study by Palomaki and colleagues of 4664 pregnancies at high-risk for Down syndrome using the MPS method had a detection rate of 98.6% with a false-positive rate of only 0.20% (3/1471) (G. E. Palomaki et al., 2011). Results also identified MPS as a successful detection method for both trisomy 18 and trisomy 13. The authors state, “Among the 99.1% of samples interpreted (1,971/1,988), observed trisomy 18 and 13 detection rates were 100% (59/59) and 91.7% (11/12) at false-positive rates of 0.28% and 0.97%, respectively... Among high-risk pregnancies, sequencing circulating cell-free DNA detects nearly all cases of Down syndrome, trisomy 18, and trisomy 13, at a low false-

positive rate. This can potentially reduce invasive diagnostic procedures and related fetal losses by 95%. Evidence supports clinical testing for these aneuploidies” (G. E. Palomaki et al., 2012).

Norton et al. (2015) reported near-perfect accuracy of detection for trisomy 21 (Down’s syndrome) with the use of cell-free DNA (cfDNA) (sensitivity, 100% [38 of 38 cases of trisomy 21]; false positive rate, 0.06% [9 false positives among 15,841 women]) in the Noninvasive Examination of Trisomy (NEXT) study. Norton and colleagues found that cfDNA testing for trisomy 21, as compared with standard screening, had a better global performance during the first trimester of pregnancy. However, they did not provide information about the 14 fetal chromosomal abnormalities in the 15,841 screened pregnancies, other than for trisomies 13, 18, and 21 (Norton et al., 2015).

Several methods for detection of fetal aneuploidy by analysis of circulating cell-free fetal DNA are commercially available. All have been validated in pregnancies deemed to be at high risk for aneuploidy. Evaluation of this technology for use in low- or average-risk pregnancies is ongoing.

A two-year longitudinal study which utilized 11,414 maternal blood samples for NIPT found that “The overall sensitivity of NIPT was 98.90, 100.00, 100.00, 90.91, 100.00, 100.00 and 100.00%, and specificities were 99.96, 99.97, 99.99, 99.96, 99.98, 100.00 and 99.99% for detecting T21, T18, T13, XO, XXX, XYY and XXY, respectively” (Garshasbi et al., 2019). Hence, this testing shows excellent potential in the detection of fetal aneuploidies.

A study out of the Illumina laboratory (formerly Verinata) compared NIPS to standard maternal serum screening in pregnant women at average risk for fetal aneuploidy. Their report included data of 5974 samples tested for trisomies 13, 18, and 21 as well as monosomy X. Aneuploidy was detected in 4.8% of samples with only 0.2% putative false-positives and 0.08% false-negatives; however, 2.8% of cases had indefinite results for a single chromosome (Futch et al., 2013). Illumina more recently reported a more extensive study consisting of 85,298 clinical cases. “Aneuploidy was detected or suspected in 2142 (2.5%) samples. For aneuploidy detected cases with known clinical outcomes, the overall positive predictive value (PPV) was 83.5% (608/728); observed PPVs for trisomies 21, 18, and 13 ranged from 50.0 to 92.8%” (Taneja et al., 2016).

Currently commercially available laboratory-developed non-invasive prenatal tests for aneuploidy include: the MaterniT21™ Plus Test (Integrated Genetics/LabCorp) (LabCorp, 2019), the verifi™ Prenatal Test (Illumina) (Illumina, 2019), the Harmony Prenatal Test™ (Ariosa Diagnostics, available from Lab Corp) (Harmony, 2019), the Panorama test (Natera, available from several reference laboratories) (Natera, 2019), the Innatal® test (Progenity, 2020), QNatal® Advanced test (QuestDiagnostics, 2019b), Prequel test (Myriad, 2020), CentoNIPT® test (Centogene, 2018), ClariTest™ Core ((GenPath, 2020), IONA® test and Sage™ prenatal screen (YourgeneHealth, 2020), Invitae NIPS test (Invitae, 2020), and Clarigo test (AgilentTechnologies, 2020).

A retrospective study by Wu et al. (2020) compared positive non-invasive prenatal screening (NIPS) results for aneuploidy to standard diagnostic tests such as traditional karyotyping and chromosomal microarray analysis (CMA). The study enrolled 551 pregnant women who screened positive for trisomy 13, trisomy 18, trisomy 21, and other sex chromosomal aneuploidies. Samples were obtained from either amniotic fluid or fetal cord blood and subsequent karyotyping or CMA confirmed a total of 256 out of 551 cases (46.4%) to possess chromosomal abnormalities concordant or partially concordant with NIPT results. Placental biopsies were obtained to assess the etiology of NIPS false positives and confined placental mosaicism (CPM) was found in 60% of the biopsies. The authors also

reported that pregnant women with advanced maternal age (>35 years) had the highest positive predictive value (PPV) for trisomy 21 (87.8%), trisomy 18 (59.3%), and trisomy 13 (37.5%), while the PPV was significantly lower for women with young maternal age (<34 years) for trisomy 21 (71.9%), trisomy 18 (0%), and trisomy 13 (16.7%). This suggests that NIPS performs better in predicting aneuploidies for pregnancies with advanced maternal age than for pregnancies with young maternal age. However, the author notes that the PPVs showed “no significant upward trend when compared based on specific age categories (an interval of 5 years), which suggested that NIPT-positive result deserves equal attention from both providers and patients regardless of maternal age” (Wu et al., 2020).

Palomaki and colleagues demonstrates that hyperglycosylated hCG (h-hCG), also known as invasive trophoblast antigen (ITA), may be a promising screening marker for Down syndrome detection in the second trimester. In the study, serum samples of 45 Down syndrome cases and 238 unaffected control pregnancies between 14 to 20 weeks of gestation were collected and measured for h-hCG, along with other screening markers (Glenn E Palomaki, Neveux, Knight, Haddow, & Pandian, 2004). As seen in the figure below, h-hCG, in combination with four other screening markers, increased the detection rate to 83% at a 5% false-positive rate from the 72% detection rate by the tripe test (Glenn E Palomaki et al., 2004; QuestDiagnostics, 2019a). In addition, “The median [h-hCG] in Down syndrome pregnancies was >3.00 multiples of the median, higher than that found for human chorionic gonadotropin (hCG).” The author recommends that “the highest screening performance for Down syndrome can be obtained by integrating first- and second-trimester serum and ultrasound markers into a single interpretation in the second trimester. This integrated test approach can detect 90% of Down syndrome pregnancies at a 3% false-positive rate” (Glenn E Palomaki et al., 2004).

Table 1. Down Syndrome Detection Rates (DRs) Obtained from Various Screening Tests¹²

Test Name	Alias	Markers Included	DR, % ^a
Triple Screen	Maternal Serum Screen 3	Age, AFP, hCG, uE3	72
Quad Screen	Maternal Serum Screen 4	Age, AFP, hCG, uE3, DIA	79
Penta Screen	Maternal Serum Screen 5	Age, AFP, hCG, uE3, DIA, h-hCG	83

^a Detection rate at a 5% false-positive rate.

V. Guidelines and Recommendations

The American College of Obstetrics and Gynecologists (ACOG) and the Society for Maternal-Fetal Medicine (SMFM) (ACOG, 2020)

ACOG and SMFM offered Recommendations for Clinical Management Guidelines for Obstetricians and Gynecologists on Screening for Aneuploidy.

The following recommendations and conclusions are based on good and consistent scientific evidence (Level A):

- “Prenatal genetic screening (serum screening with or without nuchal translucency [NT] ultrasound or cell-free DNA screening) and diagnostic testing (chorionic villus sampling [CVS] or amniocentesis) options should be discussed and offered to all pregnant women regardless of maternal age or risk of chromosomal abnormality. After review and discussion, every patient has the right to pursue or decline prenatal genetic screening and diagnostic testing.

- If screening is accepted, patients should have one prenatal screening approach, and should not have multiple screening tests performed simultaneously.
- Cell-free DNA is the most sensitive and specific screening test for the common fetal aneuploidies. Nevertheless, it has the potential for false-positive and false-negative results. Furthermore, cell-free DNA testing is not equivalent to diagnostic testing.
- All patients should be offered a second-trimester ultrasound for fetal structural defects, since these may occur with or without fetal aneuploidy; ideally this is performed between 18 and 22 weeks of gestation (with or without second-trimester maternal serum alpha-fetoprotein).
- Patients with a positive screening test result for fetal aneuploidy should undergo genetic counseling and a comprehensive ultrasound evaluation with an opportunity for diagnostic testing to confirm results.
- Patients with a negative screening test result should be made aware that this substantially decreases their risk of the targeted aneuploidy but does not ensure that the fetus is unaffected. The potential for a fetus to be affected by genetic disorders that are not evaluated by the screening or diagnostic test should also be reviewed. Even if patients have a negative screening test result, they may choose diagnostic testing later in pregnancy, particularly if additional findings become evident such as fetal anomalies identified on ultrasound examination.
- Patients whose cell-free DNA screening test results are not reported by the laboratory or are uninterpretable (a no-call test result) should be informed that test failure is associated with an increased risk of aneuploidy, receive further genetic counseling and be offered comprehensive ultrasound evaluation and diagnostic testing.
- If an enlarged nuchal translucency or an anomaly is identified on ultrasound examination, the patient should be offered genetic counseling and diagnostic testing for genetic conditions as well as a comprehensive ultrasound evaluation including detailed ultrasonography at 18–22 weeks of gestation to assess for structural abnormalities.
- Women with a positive screening test result for fetal aneuploidy should be offered further detailed counseling and testing” (ACOG, 2018, 2020).

The following recommendations and conclusions are based on limited or inconsistent scientific evidence (Level B):

- “The use of cell-free DNA screening as follow-up for patients with a screen positive serum analyte screening test result is an option for patients who want to avoid a diagnostic test. However, patients should be informed that this approach may delay definitive diagnosis and will fail to identify some fetuses with chromosomal abnormalities.
- In clinical situations of an isolated soft ultrasonographic marker (such as echogenic cardiac focus, choroid plexus cyst, pyelectasis, short humerus or femur length) where aneuploidy screening has not been performed, the patient should be counseled regarding the risk of aneuploidy associated with the finding and cell-free DNA, quad screen testing, or

amniocentesis should be offered. If aneuploidy testing is performed and is low-risk, then no further risk assessment is needed. If more than one marker is identified, then genetic counseling, maternal–fetal medicine consultation, or both are recommended.

- No method of aneuploidy screening that includes a serum sample is as accurate in twin gestations as it is in singleton pregnancies; this information should be incorporated into pretest counseling for patients with multiple gestations.
- Cell-free DNA screening can be performed in twin pregnancies. Overall, performance of screening for trisomy 21 by cell-free DNA in twin pregnancies is encouraging, but the total number of reported affected cases is small. Given the small number of affected cases it is difficult to determine an accurate detection rate for trisomy 18 and 13” (ACOG, 2018, 2020).

The following recommendations and conclusions are based primarily on consensus and expert opinion (Level C):

- “The use of multiple serum screening approaches performed independently (eg, a first-trimester screening test followed by a quad screen as an unlinked test) is not recommended because it will result in an unacceptably high positive screening rate and could deliver contradictory risk estimates.
- In multifetal gestations, if a fetal demise, vanishing twin, or anomaly is identified in one fetus, there is a significant risk of an inaccurate test result if serum-based aneuploidy screening or cell-free DNA is used. This information should be reviewed with the patient and diagnostic testing should be offered.
- Patients with unusual or multiple aneuploidies detected by cell-free DNA should be referred for genetic counseling and maternal–fetal medicine consultation” (ACOG, 2018, 2020).

ACOG also comments on the specific types of screening, which include triple, quadruple (quad) and “penta” screens. These screens may be performed in the first trimester (10-14 weeks gestation) or second trimester (15-22 weeks). Triple screens measure serum hCG [human chorionic gonadotropin], AFP [alpha-fetoprotein], and uE3 [unconjugated estriol], while the quad screen includes DIA [dimeric inhibin A] with the three previously mentioned markers. Some laboratories have been noted to offer a “penta” screen, which includes hyperglycosylated hCG along with the four analytes of the quad screen, but ACOG states that “its performance has not been evaluated rigorously in prospective studies”. ACOG discusses several testing algorithms in the include, which are summarized in the table below:

Screening Approach	Approximate Gestational Age Range for Screening (Weeks)	Detection Rate (DR) for Trisomy 21 (%)	Screen Positive Rate* (%)	Advantages	Disadvantages	Method
Cell-free DNA [†]	9–10 to term	99	2–4% Includes inability to obtain results, which is associated with increased risk [†]	1. Highest DR 2. Can be performed at any gestational age after 9–10 weeks 3. Lowest false-positive rate	Results may reflect underlying maternal aneuploidy or maternal disease	Several molecular methods
First trimester [‡]	10–13 6/7 [§]	82–87	5	1. Early screening 2. Single time point test	Lower DR than tests with first and second trimester component NT required	NT+PAPP-A, free beta hCG, +/- AFP
Quad screen [‡]	15–22	81	5	1. Single time point test 2. No specialized US required	Lower DR than first trimester and first and second trimester combined tests	hCG, AFP, uE3, DIA
Integrated [‡]	10–13 6/7 [§] , then 15–22	96	5	High DR	Two samples needed No first-trimester results NT required	NT+PAPP-A, then quad screen
Serum integrated [‡]	10–13 6/7 [§] , then 15–22	88	5	1. DR compares favorably with first-trimester screening 2. No specialized US required	Two samples needed No first-trimester results	PAPP-A + quad screen
Sequential [#] : stepwise	10–13 6/7 [§] , then 15–22	95	5	1. First-trimester results provided 2. Comparable performance to integrated, but FTS results provided First-trimester test result: Positive: diagnostic test or cell-free DNA offered Negative: no further testing Intermediate: second-trimester test offered Final: risk assessment incorporates first- and second-trimester results	Two samples needed NT required	NT+ free beta hCG + PAPP-A, +/- AFP , then quad screen NT+hCG+ PAPP-A, +/- AFP , then quad screen
Contingent screening ^{**}		88–94	5		Possibly two samples needed NT required	

Finally, ACOG notes that other trisomies, such as trisomies 16 or 22, can be tested for. However, ACOG recommends against screening for these two aneuploidies due to lack of validated data (ACOG, 2020).

Society for Maternal-Fetal Medicine (SMFM) through Choosing Wisely (SMFM, 2019)

SMFM submitted fifteen short recommendations regarding maternal and fetal medicine through Choosing Wisely. The relevant recommendation is as follows:

“Don't order serum aneuploidy screening after cfDNA [cell-free DNA] aneuploidy screening has already been performed.” (SMFM, 2019)

The National Society of Genetic Counselors (NSGC) (Devers et al., 2013)

The NSGC issued a position statement that supports noninvasive prenatal screening as an option for pregnancies considered high risk for trisomy 13, 18 or 21. “The National Society of Genetic Counselors currently supports Noninvasive Prenatal Testing/Noninvasive Prenatal Diagnosis (NIPT/NIPD) as an option for patients whose pregnancies are considered to be at an increased risk for certain chromosome abnormalities. NSGC urges that NIPT/NIPD only be offered in the context of informed consent, education, and counseling by a qualified provider, such as a certified genetic counselor. Patients whose NIPT/NIPD results are abnormal, or who have other factors suggestive of a chromosome abnormality, should receive genetic counseling and be given the option of standard confirmatory diagnostic testing” (Devers et al., 2013).

The International Society for Prenatal Diagnosis (ISPD) (Benn et al., 2013)

The ISPD recognizes that massively parallel sequencing for detection of Down syndrome can be “helpful” for women with high risk pregnancies, when “suitable genetic counseling” is provided (Benn et al., 2013).

The International Society for Prenatal Diagnosis (ISPD) (Glenn E. Palomaki et al., 2020)

The International Society for Prenatal Diagnosis released a statement regarding cell free DNA screening for Down syndrome in multifetal pregnancies. The following recommendations were made:

- “The use of first trimester cfDNA screening for the common autosomal trisomies is appropriate for twin pregnancies due to sufficient evidence showing high detection and low false positive rates with high predictive values.
- The finding of an increased risk on a cfDNA screening test in multiple pregnancies should be followed by counseling and an offer of diagnostic testing to confirm results...CVS and amniocentesis procedures in multiple pregnancies are reliable and safe when performed by a provider experienced in these situations; subsequent diagnostic tests are highly reliable... Maternal age and nuchal translucency (with or without biochemistry) detects up to 80% of Down syndrome at a 5% false positive rate in twin pregnancies.
- It is preferable for laboratories performing cfDNA testing in multifetal pregnancies to take evidence of zygosity (e.g., chorionicity, sex of the fetuses, embryo transfer history) for the interpretation of both test results and fetal fractions...Interpretation of the cfDNA test results could differ depending on test methodology, fetal fraction and chorionicity/zygosity...Fetal fractions are higher in twin pregnancies, but lower for individual fetuses when compared to singletons. Fetal fractions are correlated between dizygous twins, but can still vary two-fold.
- cfDNA based screening for common trisomies in twins provides higher positive predictive values among twin pregnancies compared with traditional serum and nuchal translucency based screening in twins, but are associated with test failures...When a cfDNA test failure

occurs consider ultrasound and diagnostic testing. If there is sufficient time, a second sample draw may also be considered.

- Screening options for triplet pregnancies are lacking and cfDNA may be a potential option. However, diagnostic testing should always be offered and the limitations of screening tests stressed” (Glenn E. Palomaki et al., 2020).

The American College of Medical Genetics and Genomics (ACMG) (Gregg et al., 2016)

The ACMG notes that “Pretest information should be provided ... to ensure patients make informed decisions. Aneuploidy screening is not a routine prenatal test; it is acceptable for patients to decline screening.” The ACMG also cautions that “All reports should clearly state that NIPS is a screening test and not diagnostic,” and that results be presented in a clear and easily understandable fashion. ACMG guidelines state that “in pregnancies with multiple gestations and/or donor oocytes, testing laboratories should be contacted regarding the validity of NIPS before it is offered to the patient as a screening option” (Gregg et al., 2016).

Practice Committee and Genetic Counseling Professional Group (GCPG) of the American Society for Reproductive Medicine (ASRM, 2020)

The American Society of Reproductive Medicine notes that screening tests (maternal serum biochemical screening, nuchal translucency and fetal anatomy scan, and noninvasive prenatal screening with cell-free DNA) cannot diagnose chromosomal aneuploidy. “In some cases, ultrasound and biochemical analytes may help identify congenital anomalies that may be associated with an aneuploid pregnancy; however, many aneuploidies (and mosaic aneuploidies in particular) do not result in visible ultrasound anomalies or skewed biochemical analytes and may be easily missed.” Cell-free DNA testing “may test for a select number of full and partial aneuploidies, or all aneuploidies within a specified chromosomal resolution, depending on the specific test used by the laboratory. If the chromosome or chromosomal segment of interest is in fact able to be assessed by the assay used, an aneuploidy may be detected. However, it is important to recognize that NIPT is not designed for the detection of mosaicism and may result in false-negative results. False-positive results may also occur because NIPT analyzes placental (and not fetal) DNA” (ASRM, 2020).

VI. State and Federal Regulations, as applicable

Fetal ultrasound uses available instrumentation and as a medical procedure is not subject to regulation by the U.S. Food and Drug Administration. Additionally, many labs have developed specific tests that they must validate and perform in house. These laboratory-developed tests (LDTs) are regulated by the Centers for Medicare and Medicaid (CMS) as high-complexity tests under the Clinical Laboratory Improvement Amendments of 1988 (CLIA '88). As an LDT, the U. S. Food and Drug Administration has not approved or cleared this test; however, FDA clearance or approval is not currently required for clinical use.

VII. Applicable CPT/HCPCS Procedure Codes

Billing applicable codes is not a guarantee of payment; see Section III for indications and limitations of coverage that may affect payment

Code Number	Code Description
81420	Fetal chromosomal aneuploidy (eg, trisomy 21, monosomy X) genomic sequence analysis panel, circulating cell-free fetal DNA in maternal blood, must include analysis of chromosomes 13, 18, and 21
81422	Fetal chromosomal microdeletion(s) genomic sequence analysis (eg, DiGeorge syndrome, Cri-du-chat syndrome), circulating cell-free fetal DNA in maternal blood
81479	Unlisted molecular pathology procedure
81508	Fetal congenital abnormalities, biochemical assays of two proteins (PAPP-A, hCG), utilizing maternal serum, algorithm reported as risk score
81509	Fetal congenital abnormalities, biochemical assays of three proteins (PAPP-A, hCG, DIA), utilizing maternal serum, algorithm reported as risk score
81510	Fetal congenital abnormalities, biochemical assays of three analytes (AFP, uE3, hCG), utilizing maternal serum, algorithm reported as risk score
81511	Fetal congenital abnormalities, biochemical assays of four analytes (AFP, uE3, hCG [any form], DIA), utilizing maternal serum, algorithm reported as a risk score (may include additional results from previous biochemical testing)
81512	Fetal congenital abnormalities, biochemical assays of four analytes (AFP, uE3, hCG [any form], DIA), utilizing maternal serum, algorithm reported as a risk score (may include additional results from previous biochemical testing)
81599	Unlisted multianalyte assay with algorithmic analysis
82105	Alpha-fetoprotein; serum
82106	Alpha-fetoprotein (AFP); amniotic fluid
82677	Estriol
84163	Pregnancy-associated plasma protein (PAPP-A)
84702	Gonadotropin, chorionic; quantitative
84703	Gonadotropin, chorionic (hCG); qualitative
84704	Gonadotropin, chorionic (hCG); free beta chain
86336	Inhibin A

88235	Tissue culture for non-neoplastic disorders; amniotic fluid or chorionic villus cells
88267	Chromosome analysis, amniotic fluid or chorionic villus, count 15 cells, 1 karyotype, with banding
88269	Chromosome analysis, in situ for amniotic fluid cells, count cells from 6-12 colonies, 1 karyotype, with banding
88271	Molecular cytogenetics, DNA probe, each (e.g., FISH)
88280	Chromosome analysis; additional karyotypes, each study
88285	Chromosome analysis; additional cells counted, each study
0168U	Fetal aneuploidy (trisomy 21, 18, and 13) DNA sequence analysis of selected regions using maternal plasma without fetal fraction cutoff, algorithm reported as a risk score for each trisomy Proprietary test: Vanadis® NIPT Lab/Manufacturer: PerkinElmer, Inc

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Procedure codes appearing in Medical Policy documents are included only as a general reference tool for each policy. They may not be all-inclusive.

Note:

1. Do not report 81508 in conjunction with 84163, 84702
2. Do not report 81509 in conjunction with 84163, 84702, 86336
3. Do not report 81510 in conjunction with 82105, 82677, 84702

VIII. Evidence-based Scientific References

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